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How Do Periodontal Infections Affect the Onset and Progression of Alzheimer's Disease?

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Abstract: Chronic infection can cause slow progressive dementia, cortical atrophy and amyloid deposition in the atrophic form of general paresis. Due to the fact that specific bacterial ligands can increase the expression of proinflammatory molecules that can activate innate and adaptive immune systems, inflammation may play a significant role in the pathogenesis of Alzheimer's disease (AD). Furthermore, there is a significant association between AD and various types of spirochete. Periodontitis is a prevalent and persistent peripheral infection that is associated with gram-negative anaerobic bacteria and is capable of showing localized and systemic inflammation and the pathogenesis of AD. In this minireview, we propose a hypothetical link between periodontitis, type 2 diabetes and AD. We also present the possible mechanistic links between periodontitis-related inflammation, type 2 diabetes and AD. Since this condition is treatable, periodontitis may be a readily-modifiable risk factor for AD.

Keywords: Periodontal disease, Alzheimer's disease, inflammation, dementia.

INTRODUCTION

Alzheimer's disease (AD) is a neurodegenerative disorder that leads to amnesia, cognitive impairment and senile dementia. AD affects personal independence, relationships and the ability to express oneself comprehensibly. The disease is a devastating and fatal condition that results in a significant burden for the individual and society [1]. For these reasons, many researchers have investigated potential treatments for AD by focusing on various risk factors and pathological theories. For the prevention and management of this chronic disease, the identification of modifiable risk factors and preventive strategies is important.

Globally, an estimated 35.6 million people live with dementia [2]. The number of people with dementia is expected to double (65.7 million) by 2030 and by 2050, the number is expected to more than triple to 115.4 million [2]. Dementia affects people in all countries, but more than half (58%) of the population with dementia lives in low- and middle-income countries [2]. The most common cause of dementia is AD [3].

Periodontal disease (PD) is an inflammatory process that involves a progressive, episodic loss of the periodontal attachment apparatus. In susceptible patients, PD ultimately results in tooth loss. In individuals younger than 35 years old, the prevalence of moderate and severe PD is less than 1%, and an increasing prevalence is reported in older age groups [4]. In the USA alone, 25% of the population that is 75 years old and older suffers from PD (moderate PD: 18%; severe PD: 7%) [4]. A higher prevalence of PD is reported in the male population, and men are more likely than women to be at risk for PD [5]. PD is also implicated as being one of the primary complications in type 2 diabetes mellitus (T2DM) [6] and is hypothesized to predispose patients to AD [7].

In the majority of cases, the cause of AD is unknown. Most experts agree that AD is just like other common, chronic conditions that are likely to develop as a result of multiple factors rather than because of a single cause. AD is pathologically and genetically linked to T2DM [8, 9], although PD is only pathologically linked to T2DM [10]. Many questions remain about whether PD or cognitive impairment came first.

PDs are recognized as infectious processes that require the presence of bacteria and a host response. Risk factors in conjunction with bacteria and the host response can affect the severity of AD, patterns of destruction and responses to therapy. In the past decade, many studies have changed the approach to studying periodontal infection and the relationship between general health and PD. In this paper, we reviewed various published articles on PD as a potentially modifiable risk factor for AD.

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PATHOLOGY OF PD

The dental plaque biofilm is the most potent cause of direct and indirect periodontal tissue damage [11]. However, the majority of periodontal tissue injury is attributed to indirect mechanisms that are initiated by dental plaque, such as aggravated inflammatory host tissue response. Multiple complex interactions occur between the host defense cells and the periodontal tissues.

The inflammatory response in PD includes the activation of leucocytes, neutrophils, T-lymphocytes and plasma cells [12]. The inflammatory response also includes the release of antibodies, lipopolysaccharides (LPSs) and chemical inflammatory mediators, such as cytokines, chemokines and C-reactive proteins. LPSs are present in the gram-negative bacterial cell walls and act as powerful stimulants for complex host responses. The initial surge of neutrophils at the site is followed by the release of cytokines by neutrophils and macrophages. The released chemical mediators include tumor necrosis factor-alpha (TNF- α), interleukin-1 (IL-1) and prostaglandins (PGs). The inflammatory process includes the stimulation of fibroblasts by IL-1 and the secretion of matrix metalloproteinases (MMPs) by polymorphonuclear neutrophils. While MMPs are responsible for increased collagen breakdown, TNF- α is primarily responsible for increased osteoclast activity that results in bone resorption. MMPs can also activate cytokines and chemokines, which can exacerbate the destructive process. The production of collagen is inhibited by reduced fibroblast activity in response to TNF- α [13-15] (Fig. 1).

Lymphocytes release antibodies as protective mechanisms. However, lymphocytes also activate the osteoclasts that result in bone loss [16]. T-lymphocytes secrete the receptor activator nuclear factor kappa-B ligand (RANKL), which is involved in osteoclast activity and leads to bone resorption. The destructive inflammatory mediators are inhibited by the presence of osteoprotegerin and the secreted tissue inhibitors of metalloproteinases (TIMPs) [14, 17, 18].

The level of periodontal destruction depends on the balance between destructive and protective inflammatory mediators. While periodontal bacteria are required for infective PD, an individual's response determines the disease progression. *In vitro*, an individual's response can be affected by genetic signaling pathways that influence the expression of inflammatory mediators in response to bacterial LPS [13-18].

During periodontitis, pathogens such as *Porphyromonas* gingivalis (P. gingivalis), Tannerella forsythia and Treponema denticola (T. denticola) are associated with the formation of biofilm [19]. In the innate immune system, a family of pattern-recognition receptors named toll-like receptors (TLRs) recognizes the pathogen-associated molecular patterns of these gram-negative bacteria [20, 21]. These receptors are predominantly expressed on the surface of neutrophils, monocytes and dendritic cells, and they orchestrate a rapid innate immune response against invading pathogens [20-22]. The activation of TLRs (except for TLR3) induces the interaction between the IL-1 receptor domain-containing protein and the adaptor protein myeloid differentiation primary response gene (MyD88). MyD88 recruits a protein kinase, such as the IL-1 receptor-associated kinase, to induce the mitogen-activated protein kinase kinase kinase (MAP3K). In turn, MAP3K activates the mitogenactivated protein kinases (MAPKs), c-Jun N-terminal kinases (JNKs), p38 and the transcription factor nuclear factor-kappa B (NF-KB) [23]. The induction of NF-KB activity contributes to the secretion of various key inflammatory mediators of chronic inflammatory conditions, including TNFa, interleukin-1b (IL-1b) and prostaglandin E2 (PGE2) [24, 25]. In patients with chronic periodontitis, enhanced levels of TNF α have been found in the serum and gingival crevicular fluid [26, 27] (Fig. 1).

In addition to the beneficial role of TLRs in the host immune response, the stimulation of TLR signalings, primarily *via* TLR2 and TLR4, may contribute to the development of a number of diseases that are promoted by chronic inflammatory processes. For example, TLR2 has



Fig. (1). Pathogenesis of periodontal diseases (PDs). LPS: lipopolysaccharide; IL: interleukin; PGs: prostaglandins; TNF: tumor necrosis factor; MMPs: metalloproteinases; RANKL: receptor activator of nuclear factor kappa-B ligand; MyD88: myeloid differentiation primary response gene: COX-2: cyclooxygenase 2; PGE2: prostaglandins E2; MAP3K: mitogen-activated protein kinase kinase kinase; JNK: c-Jun N-terminal kinases; NF-κB: nuclear factor-kappa B; TLR: toll-like receptors.

been implicated in systemic lupus erythematosus, diabetes and AD [28, 29]. Compared to periodontally healthy controls, studies have reported elevated levels of TLR2 and TLR4 in the gingival tissues of patients with chronic periodontitis [30]. In patients with PD, Lappin et al. [31] demonstrated that the levels of TLR2 and TLR4 agonists, Pam3Cys-Ser-(Lys)4 (Pam3CSK4)." and LPS are elevated in the saliva. In addition, gingival fibroblasts, which are the major cell type in the periodontal connective tissue, express both TLR2 and TLR4 [32, 33]. Gutierrez-Venegas et al. [34] demonstrated that gingival fibroblasts that are treated with the specific TLR2 ligand LPS (from the periodontitisassociated bacteria P. gingivalis) promoted the expression of cyclooxygenase-2 (COX-2) and PGE2 synthesis. While TLR2 is expressed in the gingival tissues of patients with periodontitis and the expression of the receptor increases with chronic inflammation [35], the signaling pathways that are involved in the induction of TLR2 remain unclear. Further investigation is needed to elucidate the exact role of TLR2 gene expression in PD inflammation.

LINKAGE BETWEEN PD AND T2DM (FIG. 2)

PD has been coined as the "sixth complication" of diabetes [6]. T2DM is a genetically and environmentally based chronic metabolic and vascular syndrome that is caused by insulin deficiency and alterations in lipids, carbohydrates and proteins metabolisms [10]. Hyperglycemia is the primary consequence of defects in the secretion and/or action of insulin. Alterations in insulin synthesis can affect various organs, especially including the kidneys, eyes,

In patients with PD, oral pathogens and their products can gain access to the systemic circulation, which may elicit an immune response that can disrupt the body's normal homeostasis. Combined with the presence of infection and an exaggerated host response, the accumulation of advanced glycation end products (AGEs) that occurs as a result of a chronic hyperglycemic state may explain the clinical outcomes that are observed in diabetic patients with PD [10]. Since the effective control of periodontal infection in diabetic patients reduces the level of AGEs in the serum [10], proper glycemic control is an important factor that should not be overlooked.

Bacterial products, such as endotoxin or LPS, also propagate an inflammatory response in the host through the TLRs, which induces an inflammatory cascade [36]. These receptors play an important role in the innate immune response, particularly in the initial interaction between the infecting microorganisms, such as *P. gingivalis* and phagocytic cells of the monocyte lineage [37]. Genetic and biochemical studies have shown that the toll protein family members play a critical role in the immediate response to infection [38, 39]. While LPS monocyte interactions provide one of the best-studied models of innate immunity using gram-negative bacteria and the bacterial endotoxin, the mechanisms behind PD and the regulation of TLR protein expressions are not well understood. Collectively, these



Fig. (2). Schematics of the linkages among periodontal disease; Alzheimer's disease and type 2 diabetes mellitus. LPS: lipopolysaccharide; IL: interleukin; PGs: prostaglandins; TNF: tumor necrosis factor; MMPs: metalloproteinases; RANKL: receptor activator of nuclear factor kappa-B ligand; TLR: toll-like receptors; AGE: advanced glycation end products; TLPs: toll-like receptor proteins; RAGE: receptor of advanced glycation end products.

studies have provided insight into the molecular mechanisms that support the observed epidemiological associations between PD and diabetes.

LINKAGE BETWEEN T2DM AND AD (FIG. 2)

In addition to having neuroprotective effects, a recent study suggests that insulin plays primary roles in reducing blood sugar levels and acting as a growth factor in neuronal stem cell activation [40]. Independent of cerebrovascular disease, hyperinsulinemia is a plausible risk factor for lateonset Alzheimer's disease (LOAD), as insulin can cross the blood-brain barrier [41]. Moreover, peripheral insulin infusion may affect amyloid 642 (A642) levels in cerebrospinal fluid in the elderly [42]. A β 42 is a marker for amyloid β (A β) clearance in the brain, and A β 42 is an indirect marker for the risk of LOAD. Furthermore, the number of insulin receptors in the brain, especially in the structures in the hippocampus and entorhinal cortex that are affected early during LOAD [43, 44] can be increased. The insulin-degrading enzyme (IDE) also plays a role in the clearance of AB in the brain, as both insulin and AB compete for IDE [45]. In the pathogenesis of LOAD, the deposition of key markers (A β and tau protein phosphorylation) can be increased by the presence of insulin in brain [41].

The pathways related to insulin in the periphery with $A\beta$ clearance in the brain are multiple and complex. Craft el al. reviewed the literature on how peripheral hyperinsulinemia affects amyloid beta clearance in the brain [8]. Due to saturation above physiologic levels, one potential pathway is that peripheral hyperinsulinemia downregulates insulin uptake in the blood brain barrier [18]. Peripheral hyperinsulinemia may result in the reduction of 1) insulin levels in the brain and the downregulation of IDE expression [46] and 2) IDE-mediated amyloid clearance [45]. This complex observation supports the seemingly paradoxical use of the insulin sensitizer rosiglitazone [9, 47] and intranasal insulin [48] in the treatment of LOAD.

Elevated glucose concentration promotes the accrual of AGEs, AGEs are closely linked with both glycemia and diabetes. In hyperglycemic environments, diabetic animal and human tissues contain increased AGE levels that promote the up-regulation of the AGE receptor (RAGE) [49]. AGE is associated with the traditional microvascular complications of T2DM [50]. Increased expressions of RAGE are also observed in LOAD [51], and the expression of RAGE is enhanced in blood vessels that are adjacent to A β deposits in the LOAD brain [52].

LINKAGE BETWEEN PD AND AD (FIG. 2)

Due to the interaction between periodontopathic bacteria and the host response, a significant number of inflammatory molecules (IL-1 β , IL-6 and TNF- α) are produced during brain inflammation [53]. In the brains of AD patients compared to non-AD subjects, studies have reported a relatively higher spirochetes burden, including *Treponema* species (periodontal pathogens) [54, 55]. Thus, inflammatory molecules and the aforementioned pathogens may play a role in the brain inflammation that characterizes and affects the expression of AD [56]. Moreover, clinical studies have reported that tooth loss is a significant risk factor for AD and/or dementia [57, 58]. While tooth loss may have several causes [59, 60], PD is one of the major causative factors for loss of tooth in adults. Even after extensive adjustments for confounders, the Third National Health and Nutrition Examination Survey (NHANES III) reported that gingival bleeding, loss of periodontal attachment and serum *P. gingivalis* IgG, all of which can indicate PD, were significantly associated with lower cognitive function [61, 62].

Inflammation is one of the key features of AD and provides the foundation for the hypothesis that inflammatory and infectious conditions may be risk factors for AD. PD can contribute to the expression of AD via several mechanisms [56]. Peripheral inflammation and infections play important roles in the pathogenesis of AD [63-65] and affect oral bacteria [66-69]. Furthermore, the systemic dissemination of bacterial species from local infections may affect AD [68]. During various surgical and non-surgical procedures, oral bacteria from the periodontal species can gain access to systemic circulation and colonize distant anatomical sites, which can lead to pathologies such as endocarditis and brain and lung abscesses [70]. While they were absent in healthy controls, spirochetes have been found in the blood, cerebrospinal fluid and brain samples of AD patients [54, 71], which further supports the colonization occurring via systemic spread. Using molecular techniques with Treponema species, including T. denticola, a pathogenic periodontal bacterium was detected in the brains of AD patients [55]. T. denticola belongs to the same class as T. pallidum and is known to induce pathologic conditions that are comparable with the pathology found in AD [68].

Among periodontal patients, a significant number of IgG antibodies to *T. denticola* have been reported [72]. However, no study has investigated the levels of IgG antibodies among AD patients, and there are no clinical studies that directly link PD and AD. Some studies have used antibodies to periodontal bacteria to discover associations between PDs and other systemic diseases [73-75]. The elevated antibodies *Actinobacillus actinomycetemcomitans* and *P. gingivalis* are better associated with cardiovascular disease than with clinical PD measures [76] which suggest that antibodies to periodontopathic species may provide an index of exposure for investigations of the relationship between PD and systemic diseases.

The biological reason for an association between antibody levels to periodontal bacteria and AD is unknown. AD subjects have been shown to have poor oral hygiene [77], which would suggest higher antibody levels and increased bacterial colonization. Over 500 bacterial species can colonize the supragingival and subgingival environments (the areas above and below the gingival margin, respectively) around the teeth [11]. The presence of pathogenic species has also been reported in healthy subjects. Recent studies suggest that the host inflammatory response may be more important in bacterial biofilm formation than was originally thought [78]. Furthermore, recent studies on the association between measures of periodontal infections and cognitive functions in the normal population have suggested that the association may be established even before the onset of AD [61, 62].

One study has indicated that subjects with TNF- α and IgG antibodies against periodontal pathogens have higher risk for AD, which suggests that TNF- α may not be related to the number of positive antibody responses [79]. In the presence of systemic inflammation, periodontal infection may lead to an increased risk for systemic diseases. Increased age and the presence of teeth may influence antibody levels [80], and the elderly may also have a reduced antibody response [81, 82]. Since AD patients tend to be older than non-AD patients, the level of antibody response in AD may actually be underestimated. Furthermore, the antibody response reflects the host immune function, so antibody titres to periodontal bacteria may compliment plasma TNF- α levels and improve the clinical diagnosis of AD. In this way, AD patients can be differentiated from cognitively normal subjects.

CONCLUSION

Patients with no cognitive impairment who have elevated antibodies to PD bacteria may have an increased risk of AD onset and/or progression. At the same time, the incidence and severity of T2DM from PD may aggravate cognitive impairment. Inflammatory chemical mediators that exist in the presence of bacterial products and alterations in the inflammatory gene expression can contribute to AD. The goals of this study were to understand the association between chronic oral periodontal infection and AD and to determine whether the chronic oral infection could contribute to the risk of AD expression. The current findings suggest an increased risk; however, additional cohort studies that profile oral clinical presentation with systemic response and AD are needed, and prospective studies can evaluate any cause-andeffect association. The hope is that future studies will elucidate the importance of maintaining oral health as a fundamental part of healthy aging and to lower the risk of these types of neurological changes. Global attention and action are needed to support this emerging field of research. In the future, dementia may be prevented by combining antibiotic, antiviral and anti-inflammatory therapies.

LIST OF ABBREVIATIONS

AD	=	Alzheimer's disease
Αβ42	=	Amyloid β42
IDE	=	Insulin-degrading enzyme
LOAD	=	Late-onset AD
LPS	=	Lipopolysaccharides
MMPs	=	Matrix metalloproteinases
PD	=	Periodontal disease
TLRs	=	Toll-like receptors
TNF-α	=	Tumor necrosis factor-alpha
T2DM	=	Type 2 diabetes mellitus

CONFLICT OF INTEREST

The authors declare that there are no conflicts of interest.

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REFERENCES

- Richards SS, Hendrie HC. Diagnosis, management, and treatment of Alzheimer disease: a guide for the internist. Arch Int Med 1999; 159: 789-98.
- [2] Dementia cases set to triple by 2050 but still largely ignored; Available from http://www.who.int/mediacentre/news/releases/2012/dementia_201 20411/en/ [Accessed on 16-06-2012).
- [3] World Health Organisation Dementia; Available from http://www.who.int/mediacentre/factsheets/fs362/en/index.html
 [Accessed on 16-06-2012].
- [4] Eke P, Barker L. Prevalence of Periodontal Disease in the United States: NHANES 1999-2004. International Association of Dental Research; 2007.
- [5] Shiau HJ, Reynolds MA. Sex differences in destructive periodontal disease: exploring the biologic basis. J Periodontol 2010: 81(11): 1505-17.
- [6] Rajhans NS, Kohad RM, Chaudhari VG, Mhaske NH. A clinical study of the relationship between diabetes mellitus and periodontal disease. J Indian Soc Periodontol 2011; 15(4): 388-92.
- [7] Sparks Stein P, Steffen MJ, Smith C, et al. Serum antibodies to periodontal pathogens are a risk factor for Alzheimer's disease. Alzheimers Dement 2012; 8(3): 196-203.
- [8] Craft S. Insulin resistance and Alzheimer's disease pathogenesis: potential mechanisms and implications for treatment. Curr Alzheimer Res 2007; 4(2): 147-52.
- [9] Risner ME, Saunders AM, Altman JF, et al. Efficacy of rosiglitazone in a genetically defined population with mild-to-moderate Alzheimer's disease. Pharmacogenom J 2006; 6(4): 246-54.
- [10] Southerland JH, Taylor GW, Offenbacher S. Diabetes and periodontal infection: making the connection. Clin Diabetes 2005; 23(4): 171-8.
- [11] Haffajee AD, Socransky SS. Introduction to microbial aspects of periodontal biofilm communities, development and treatment. Periodontology 2006; 42: 7-12.
- [12] Tatakis DN, Kumar PS. Etiology and pathogenesis of periodontal diseases. Dent Clin North Am 2005; 49(3): 491-516.
- [13] Graves DT, Cochran D. The contribution of interleukin-1 and tumor necrosis factor to periodontal tissue destruction. J Periodontol 2003; 74(3): 391-401.
- [14] Kinney JS, Ramseier CA, Giannobile WV. Oral fluid-based biomarkers of alveolar bone loss in periodontitis. Ann NY Acad Sci 2007; 1098: 230-51.
- [15] Mark LL, Haffajee AD, Socransky SS, *et al.* Effect of the interleukin-1 genotype on monocyte IL-1beta expression in subjects with adult periodontitis. J Periodontal Res 2000; 35(3): 172-7.
- [16] Manolagas SC. Estrogen loss, cytokines, macrophages, lymphocytes, osteoclasts and bone loss: Six characters in search of an author or an endocrine-immune system relay causing osteoporosis? BoneKEy-Osteovision 2002; 2002: 1138.
- [17] Taubman MA, Valverde P, Han X, Kawai T. Immune response: the key to bone resorption in periodontal disease. J Periodontol 2005; 76(11 Suppl): 2033-41.
- [18] Barksby HE, Nile CJ, Jaedicke KM, Taylor JJ, Preshaw PM. Differential expression of immunoregulatory genes in monocytes in response to Porphyromonas gingivalis and Escherichia coli lipopolysaccharide. Clin Exp Immunol 2009; 156(3): 479-87.

- [19] Pihlstrom BL, Michalowicz BS, Johnson NW. Periodontal diseases. Lancet 2005; 366(9499): 1809-20.
- [20] Akira S, Takeda K, Kaisho T. Toll-like receptors: critical proteins linking innate and acquired immunity. Nat Immunol 2001; 2(8): 675-80.
- [21] Kumar H, Kawai T, Akira S. Toll-like receptors and innate immunity. Biochem Biophys Res Commun 2009; 388(4): 621-5.
- [22] Sun Y, Shu R, Li CL, Zhang MZ Gram-negative periodontal bacteria induce the activation of Toll-like receptors 2 and 4, and cytokine production in human periodontal ligament cells. J Periodontol 2010; 81(10): 1488-96.
- [23] O'Neill LA. Therapeutic targeting of Toll-like receptors for inflammatory and infectious diseases. Curr Opin Pharmacol 2003; 3(4): 396-403.
- [24] Carmody RJ, Chen YH. Nuclear factor-kappaB: activation and regulation during toll-like receptor signaling. Cell Mol Immunol 2007; 4(1): 31-41.
- [25] Wang P, Meinhardt B, Andre R, et al. The interleukin-1-related cytokine IL-1F8 is expressed in glial cells, but fails to induce IL-1beta signalling responses. Cytokine 2005; 29(6): 245-50.
- [26] Passoja A, Puijola I, Knuuttila M, et al. Serum levels of interleukin-10 and tumour necrosis factor-alpha in chronic periodontitis. J Clin Periodontol 2010; 37(10): 881-7.
- [27] Sakai A, Ohshima M, Sugano N, Otsuka K, Ito K. Profiling the cytokines in gingival crevicular fluid using a cytokine antibody array. J Periodontol 2006; 77(5): 856-64.
- [28] Chen K, Huang J, Gong W, et al. Toll-like receptors in inflammation, infection and cancer. Int Immunopharmacol 2007; 7(10): 1271-85.
- [29] Urbonaviciute V, Furnrohr BG, Meister S, et al. Induction of inflammatory and immune responses by HMGB1-nucleosome complexes: implications for the pathogenesis of SLE. J Exp Med 2008; 205(13): 3007-18.
- [30] Sun Y, Guo QM, Liu DL, Zhang MZ, Shu R. *In vivo* expression of Toll-like receptor 2, Toll-like receptor 4, CSF2 and LY64 in Chinese chronic periodontitis patients. Oral Dis 2010; 16(4): 343-50.
- [31] Lappin DF, Sherrabeh S, Erridge C. Stimulants of Toll-like receptors 2 and 4 are elevated in saliva of periodontitis patients compared with healthy subjects. J Clin Periodontol 2011; 38(4): 318-25.
- [32] Wang PL, Azuma Y, Shinohara M, Ohura K. Toll-like receptor 4mediated signal pathway induced by Porphyromonas gingivalis lipopolysaccharide in human gingival fibroblasts. Biochem Biophys Res Commun 2000; 273(3): 1161-7.
- [33] Tabeta K, Yamazaki K, Akashi S, et al. Toll-like receptors confer responsiveness to lipopolysaccharide from Porphyromonas gingivalis in human gingival fibroblasts. Infect Immun 2000; 68(6): 3731-5.
- [34] Gutierrez-Venegas G, Cruz-Arrieta S, Villeda-Navarro M, Mendez-Mejia JA. Histamine promotes the expression of receptors TLR2 and TLR4 and amplifies sensitivity to lipopolysaccharide and lipoteichoic acid treatment in human gingival fibroblasts. Cell Biol Int 2011; 35(10): 1009-17.
- [35] Mori Y, Yoshimura A, Ukai T, *et al.* Immunohistochemical localization of Toll-like receptors 2 and 4 in gingival tissue from patients with periodontitis. Oral Microbiol Immunol 2003; 18(1): 54-8.
- [36] Wittebole X, Coyle SM, Kumar A, et al. Expression of tumour necrosis factor receptor and Toll-like receptor 2 and 4 on peripheral blood leucocytes of human volunteers after endotoxin challenge: a comparison of flow cytometric light scatter and immunofluorescence gating. Clin Exp Immunol 2005; 141(1): 99-106.
- [37] Kirschning CJ, Wesche H, Merrill Ayres T, Rothe M. Human tolllike receptor 2 confers responsiveness to bacterial lipopolysaccharide. J Exp Med 1998; 188(11): 2091-7.
- [38] Takeda K, Kaisho T, Akira S. Toll-like receptors. Annu Rev Immunol 2003; 21: 335-76.
- [39] Akira S, Sato S. Toll-like receptors and their signaling mechanisms. Scand J Infect Dis 2003; 35(9): 555-62.
- [40] Li L, Holscher C. Common pathological processes in Alzheimer disease and type 2 diabetes: a review. Brain Res Rev 2007; 56(2): 384-402.
- [41] Park CR. Cognitive effects of insulin in the central nervous system. Neurosci Biobehav Rev 2001; 25(4): 311-23.

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- [42] Watson GS, Peskind ER, Asthana S, et al. Insulin increases CSF Abeta42 levels in normal older adults. Neurology 2003; 60(12): 1899-903.
- [43] Small SA, Perera GM, DeLaPaz R, Mayeux R, Stern Y. Differential regional dysfunction of the hippocampal formation among elderly with memory decline and Alzheimer's disease. Ann Neurol 1999; 45(4): 466-72.
- [44] Frolich L, Blum-Degen D, Bernstein HG, et al. Brain insulin and insulin receptors in aging and sporadic Alzheimer's disease. J Neural Transm 1998; 105(4-5): 423-38.
- [45] Farris W, Mansourian S, Chang Y, et al. Insulin-degrading enzyme regulates the levels of insulin, amyloid beta-protein, and the betaamyloid precursor protein intracellular domain *in vivo*. Proc Natl Acad Sci USA 2003; 100(7): 4162-7.
- [46] Zhao L, Teter B, Morihara T, et al. Insulin-degrading enzyme as a downstream target of insulin receptor signaling cascade: implications for Alzheimer's disease intervention. J Neurosci 2004; 24(49): 11120-6.
- [47] Watson GS, Cholerton BA, Reger MA, et al. Preserved cognition in patients with early Alzheimer disease and amnestic mild cognitive impairment during treatment with rosiglitazone: a preliminary study. Am J Geriatr Psychiatry 2005; 13(11): 950-8.
- [48] Reger MA, Watson GS, Frey WH, et al. Effects of intranasal insulin on cognition in memory-impaired older adults: modulation by APOE genotype. Neurobiol Aging 2006; 27(3): 451-8.
- [49] Negrean M, Stirban A, Stratmann B, et al. Effects of low- and high-advanced glycation endproduct meals on macro- and microvascular endothelial function and oxidative stress in patients with type 2 diabetes mellitus. Am J Clin Nutr 2007; 85(5): 1236-43.
- [50] Sullivan KA, Feldman EL. New developments in diabetic neuropathy. Curr Opin Neurol 2005; 18(5): 586-90.
- [51] Lue LF, Walker DG, Brachova L, et al. Involvement of microglial receptor for advanced glycation endproducts (RAGE) in Alzheimer's disease: identification of a cellular activation mechanism. Exp Neurol 2001; 171(1): 29-45.
- [52] Deane R, Yan DS, Submamaryan RK, *et al.* RAGE mediates amyloid-beta peptide transport across the blood-brain barrier and accumulation in brain. Nat Med 2003; 9(7): 907-13.
- [53] D'Aiuto F, Parkar M, Andreou G, et al. Periodontitis and systemic inflammation: control of the local infection is associated with a reduction in serum inflammatory markers. J Dent Res 2004; 83(2): 156-60.
- [54] Miklossy J. Alzheimer's disease--a spirochetosis? Neuroreport 1993; 4(9): 1069.
- [55] Riviere GR, Riviere KH, Smith KS. Molecular and immunological evidence of oral Treponema in the human brain and their association with Alzheimer's disease. Oral Microbiol Immunol 2002; 17(2): 113-8.
- [56] Kamer AR, Craig RG, Dasanayake AP, et al. Inflammation and Alzheimer's disease: possible role of periodontal diseases. Alzheimers Dement 2008; 4(4): 242-50.
- [57] Gatz M, Mortimer JA, Fratiglioni L, et al. Potentially modifiable risk factors for dementia in identical twins. Alzheimers Dement 2006; 2(2): 110-7.
- [58] Stein PS, Desrosiers M, Donegan SJ, Yepes JF, Kryscio RJ. Tooth loss, dementia and neuropathology in the Nun study. J Am Dent Assoc 2007; 138(10): 1314-22.
- [59] Al-Shammari KF, Al-Khabbaz AK, Al-Ansari JM, Neiva R, Wang HL. Risk indicators for tooth loss due to periodontal disease. J Periodontol 2005; 76(11): 1910-8.
- [60] Jovino-Silveira RC, Caldas Ade F, de Souza EH, Gusmao ES. Primary reason for tooth extraction in a Brazilian adult population. Oral Health Prev Dent 2005; 3(3): 151-7.
- [61] Noble JM, Borrell LN, Papapanou PN, et al. Periodontitis is associated with cognitive impairment among older adults: analysis of NHANES-III. J Neurol Neurosurg Psychiatry 2009; 80(11): 1206-11.
- [62] Stewart R, Sabbah W, Tsakos G, D'Aiuto F, Watt RG. Oral health and cognitive function in the Third National Health and Nutrition Examination Survey (NHANES III). Psychosom Med 2008; 70(8): 936-41.
- [63] Holmes C, El-Okl M, Williams AL, et al. Systemic infection, interleukin 1beta, and cognitive decline in Alzheimer's disease. J Neurol Neurosurg Psychiatry 2003; 74(6): 788-9.

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- [64] Lerner AJ, Hedera P, Koss E, Stuckey J, Friedland RP. Delirium in Alzheimer disease. Alzheimer Dis Assoc Disord 1997; 11(1): 16-20.
- [65] Schmidt R, Schmidt H, Curb JD, et al. Early inflammation and dementia: a 25-year follow-up of the Honolulu-Asia Aging Study. Ann Neurol 2002; 52(2): 168-74.
- [66] Balin BJ, Little CS, Hammond CJ, et al. Chlamydophila pneumoniae and the etiology of late-onset Alzheimer's disease. J Alzheimers Dis 2008; 13(4): 371-80.
- [67] Itzhaki RF, Wozniak MA. Herpes simplex virus type 1 in Alzheimer's disease: the enemy within. J Alzheimers Dis 2008; 13(4): 393-405.
- [68] Miklossy J. Chronic inflammation and amyloidogenesis in Alzheimer's disease - role of Spirochetes. J Alzheimers Dis 2008; 13(4): 381-91.
- [69] Urosevic N, Martins RN. Infection and Alzheimer's disease: the APOE epsilon4 connection and lipid metabolism. J Alzheimers Dis 2008; 13(4): 421-35.
- [70] Zijlstra EE, Swart GR, Godfroy FJ, Degener JE. Pericarditis, pneumonia and brain abscess due to a combined Actinomyces-Actinobacillus actinomycetemcomitans infection. J Infect 1992; 25(1): 83-7.
- [71] Miklossy J, Kasas S, Janzer RC, Ardizzoni F, Van der Loos H. Further ultrastructural evidence that spirochaetes may play a role in the aetiology of Alzheimer's disease. Neuroreport 1994; 5(10): 1201-4.
- [72] Socransky SS, Haffajee AD, Cugini MA, Smith C, Kent RL. Microbial complexes in subgingival plaque. J Clin Periodontol 1998; 25(2): 134-44.
- [73] Beck JD, Offenbacher S. Systemic effects of periodontitis: epidemiology of periodontal disease and cardiovascular disease. J Periodontol 2005; 76(11 Suppl): 2089-100.

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- [74] Kshirsagar AV, Offenbacher S, Moss KL, Barros SP, Beck JD. Antibodies to periodontal organisms are associated with decreased kidney function. The Dental Atherosclerosis Risk In Communities study. Blood Purif 2007; 25(1): 125-32.
- [75] Pussinen PJ, Paju S, Mantyla P, Sorsa T. Serum microbial- and host-derived markers of periodontal diseases: a review. Curr Med Chem 2007; 14(22): 2402-12.
- [76] Mustapha IZ, Debrey S, Oladubu M, Ugarte R. Markers of systemic bacterial exposure in periodontal disease and cardiovascular disease risk: a systematic review and meta-analysis. J Periodontol 2007; 78(12): 2289-302.
- [77] Ship JA. Oral health of patients with Alzheimer's disease. J Am Dent Assoc 1992; 123(1): 53-8.
- [78] Van Dyke TE, Kornman KS. Inflammation and factors that may regulate inflammatory response. J Periodontol 2008; 79(8 Suppl): 1503-7.
- [79] Kamer AR, Craig RG, Pirraglia E, et al. TNF-alpha and antibodies to periodontal bacteria discriminate between Alzheimer's disease patients and normal subjects. J Neuroimmunol 2009; 216(1-2): 92-7.
- [80] Papapanou PN, Neiderud AM, Disick E, et al. Longitudinal stability of serum immunoglobulin G responses to periodontal bacteria. J Clin Periodontol 2004; 31(11): 985-90.
- [81] McArthur WP. Effect of aging on immunocompetent and inflammatory cells. Periodontology 2000; 16: 53-79.
- [82] Papapanou PN, Neiderud AM, Papadimitriou A, Sandros J, Dahlen G. "Checkerboard" assessments of periodontal microbiota and serum antibody responses: a case-control study. J Periodontol 2000; 71(6): 885-97.